REVIEW

Open Access

Clarithromycin-resistant *Helicobacter pylori* in Africa: a systematic review and metaanalysis

Komla Mawunyo Dossouvi^{1*}, Tchilabalo Bouyo², Simon Sognonnou³, Ephraim Ehidiamen Ibadin⁴, Lu-chao Lv⁵, Bissoume Sambe Ba⁶, Abdoulaye Seck⁷, Sika Dossim⁸, Fábio Parra Sellera^{9,10}, Makhtar Camara¹¹, Amr El Kelish^{12,13} and Stella Ifeanyi Smith^{14,15}

Abstract

Background In 2022, approximately 56.5% of adults and 47.1% of children and adolescents were affected by *Helicobacter pylori* (*H. pylori*) infection in Africa, and clarithromycin-resistant *H. pylori* (CRHp) strains have become global priority pathogens. Therefore, this study aimed to conduct the first comprehensive systematic review and meta-analysis of CRHp in Africa.

Methods This investigation was conducted according to the guidelines of the Preferred Reporting Items for Systematic reviews and Meta-Analyses (The PRISMA 2020). Literature search of electronic databases (Google Scholar, African Journals Online, ResearchGate, PubMed, Embase, and Scopus) was performed using keywords "clarithromycin", "*Helicobacter pylori*", "African country name", "mutation in the 23S rRNA".

Results Sixty-five studies involving 5,313 *H. pylori* strains isolated over 26 years (1997–2022) from 23 African countries were included in this study. The samples from which CRHp was isolated included gastric biopsy (60/63; 95%), and stool (4/63; 6%). The pooled prevalence of CRHp in Africa was 27% (95% Cl: 22, 33). There was a steady trend in the prevalence of CRHp isolated in Africa over the 26 years ($R^2 = 0.0001$, p = 0.92, slope coefficient of -0.05x). Ten types of 23S rRNA mutations (conferring clarithromycin resistance) were identified, and included mainly A2143G (465 *H. pylori* strains out of 1178 tested) and A2142G (344 *H. pylori* strains out of 1027).

Conclusion To enhance the accuracy and validity of surveillance data for *H. pylori* in Africa, there is an urgent need for implementing standardized microbiological methods for resistance detection. The prevalence of CRHp reported in this study was very similar to the overall global prevalence and there is a need for more representative studies on CRHp in Africa. While waiting for this, the treatment of *H. pylori* infections must be based on the guidelines of the AHMSG first Lagos consensus.

Keywords *Helicobacter pylori*, Clarithromycin resistance, Antimicrobial resistance in Africa, Global priority pathogens, Systematic review, Meta-analysis

*Correspondence: Komla Mawunyo Dossouvi dossouvikomlamawunyo@gmail.com

Full list of author information is available at the end of the article

© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article are included in the article's Creative Commons licence, unless indicate otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creative.commons.org/licenses/by-nc-nd/4.0/.

Introduction

Helicobacter pylori, (H. pylori) is responsible for one of the most common human bacterial infections worldwide [1, 2]. In 2022, Africa had the highest prevalence of H. pylori infection compared to other global regions [2]. The prevalence of H. pylori infection in Africa (56.5% for adults and 47.1% for children and adolescents) may be underestimated because of the scarcity of data on asymptomatic H. pylori carriage in most African countries [2, 3]. H. pylori infections are usually associated with poor sanitation and unclean water supplies [4].

H. pylori is also responsible for approximately 90% of the global burden of non-cardiac gastric cancer [5]. In 2020, the International Agency for Research on Cancer has classified *H. pylori* as a class 1 carcinogen, with a higher incidence of cancer than the human papillomavirus, hepatitis B virus, and hepatitis C virus [6]. To establish and maintain infections, *H. pylori* strains use several virulence factors such as adhesins (*babA*), urease (*ure* operon), vacuolating cytotoxin (*vacA*), immunodominant antigen (*cagA*), and the induced by contact with epithelium (*iceA*), which are most frequently associated with peptic ulceration and increased production of IL-8 [7, 8].

Advances in medicine have improved the treatment of *H. pylori* infection. In Africa, first-line therapy combines antibiotics (amoxicillin, clarithromycin) with proton pump inhibitors (PPIs), whereas second-line or salvage therapies (levofloxacin-based triple therapy, sequential non-bismuth quadruple therapy, or bismuth-based quadruple therapy) combine antibiotics (amoxicillin, clarithromycin, nitroimidazole, levofloxacin, and tetracycline) with PPIs and bismuth compounds [3].

The antimicrobial resistance has not spared *H. pylori* strains, with global resistance rates of 24% for levofloxacin, 34% for clarithromycin, and 55% for metronidazole [9]. Clarithromycin inhibits *H. pylori* protein synthesis by interacting with the peptidyl transferase ring in the V region of the 23S ribosomal RNA (rRNA) subunit of *H. pylori* strains. Mutations in the 23S rRNA are responsible for clarithromycin resistance [10–12]. The resistance of *H. pylori* to clarithromycin is associated with a sevenfold risk of treatment failure when using a clarithromycin-based regimen [9]. Therefore, in areas with low clarithromycin resistance (<15%), the treatment algorithm consists of amoxicillin, clarithromycin, and a PPI, whereas second-line therapeutic regimens are used for areas with clarithromycin resistance > 15% [13].

In 2017, the World Health Organization (WHO) released a global priority list of antibiotic-resistant bacteria, and clarithromycin-resistant *H. pylori* (CRHp) strains were ranked as high-priority pathogens for which new antibiotics are urgently needed [14]. Therefore, from an epidemiological standpoint, monitoring the spread of CRHp has become a priority for global public health authorities.

The previous pooled prevalences of CRHp determined in Africa should be taken cautiously since the studies were only limited to a few African countries [9, 15]. To address the lack of quality data on CRHp in Africa, this study aimed to conduct the first comprehensive systematic review and meta-analysis of CRHp in Africa.

Methods

This systematic review and meta-analysis was conducted according to the guidelines of the Preferred Reporting Items for Systematic reviews and Meta-Analyses (The PRISMA 2020) [16].

Literature review

A comprehensive literature search of electronic databases (Google Scholar, African Journals Online, ResearchGate, PubMed, Embase, and Scopus) was performed using keywords "clarithromycin", "*Helicobacter pylori*", "Africa", "African country name", "mutation in the 23S rRNA"). The database search was conducted from March 23, 2024, to April 30, 2024, and the studies written in French and English were included in this systematic review and meta-analysis.

Eligibility criteria

This systematic review and meta-analysis included original peer-reviewed research articles and theses reporting CRHp in African countries. Studies reporting CRHp using both genotypic and phenotypic methods, were included. After identification, duplicate articles were excluded. Reviews, commentaries, perspectives, and non-peer-reviewed articles, reporting CRHp in Africa were also excluded. Furthermore, peer-reviewed research articles and theses on *H. pylori* strains isolated from outside Africa were also excluded.

Quality assessment

Two authors performed the article quality assessment using the Joanna Briggs Institute (JBI) Prevalence Critical Appraisal Tool [17]. The results of the article quality assessment are presented in the Additional file 1. The JBI prevalence appraisal tool includes 10 questions for each article to be answered Yes (Y) or No (N). A positive answer (Y) was worth 10%, and the total number of points that could be obtained for an article was 100%. Studies with a score of \geq 50% were considered of good quality and were included in the analyses (Additional file 1).

Data extraction

For each study included in this systematic review and meta-analysis, the following data were extracted: country, authors and references, sample collection period, study design, sample from which the *H. pylori* strains were isolated, methods used to assess clarithromycin resistance, total number of *H. pylori* strains studied, number of CRHp strains, mutations within the 23S rRNA conferring clarithromycin resistance and virulence genes. Two authors conducted the data extraction, and disagreements were resolved by discussion, data cross-checking, and validation.

Data analysis

The pooled prevalences and their *p*-values, forest plots and their *p*-values, funnel plot, and meta-regression were obtained using Stata v17.0 with commands such as *'metaprop'*, *'metafunnel'*, *'metabias'*, and *'meta regress'*. The pooled prevalence were presented with a 95% CI, corresponding *p*-value and forest plot. Funnel plot symmetry and Egger's test statistics were used to evaluate prevalence publication bias [18, 19]. Microsoft Excel 2016 v2.0, was used to perform the remaining statistical analyses, and draw associated graphs. The *p*-values obtained using Microsoft Excel were calculated based on the chi-square proportion comparison test. The level of significance for all statistical tests was set at *p* < 0.05.

Selection of studies

A literature search of public databases (Google Scholar, ResearchGate, African Journals Online, PubMed, Embase, and Scopus), generated 923 studies. Subsequently, 426, 327, 19, 46, and 40 studies were excluded for duplication, data outside Africa, other types of studies (reviews, commentaries, non-peer-reviewed articles and perspective articles), studies reporting bacteria other than *H. pylori*, and studies without any data on clarithromycin resistance, respectively. The remaining 65 studies (64 research articles and one thesis) were included in this systematic review and meta-analysis (Fig. 1). Table 1 shows the data collected from the 65 studies.

Results

Characteristics of included studies

The article quality assessment using the JBI prevalence critical appraisal tool provided an overall risk of bias assessment score of 89.5% (Additional file 1). A total of 5,313 *H. pylori* strains were studied across 65 studies and were isolated over 26 years (1997–2022) from 23 African countries (Fig. 2). Thirty-two studies mentioned the type of study, including 18 (56%) cross-sectional studies, 10 (31%) prospective studies, three (9%) observational studies, and one (3%) case-control study. The number of studies on CRHp per year has increased over the years in Africa (Fig. S1).

Samples carrying CRHp

Sixty-three studies (97%) specified the samples from which CRHp was isolated, including gastric biopsy (60/63; 95%), and stool (4/63; 6%).

Methods used to assess clarithromycin resistance

Sixty-two studies (95%) specified the methods used to assess clarithromycin resistance. The phenotypic methods (48/62; 77%), included the Kirby-Bauer disc diffusion method (22/62; 36%), E-test (20/62; 32%), and agar dilution method (6/62; 10%). Genotypic methods (38/62; 61%), included real-time polymerase chain reaction (RT-PCR) (17/62; 27%), polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) (9/62; 15%), end-point PCR (5/62; 8%), DNA sequencing (5/62; 8%), and the Genotype *Helicobacter* DR Kit (2/62; 3%).

Pooled prevalence of CRHp

A total of 5,313 *H. pylori* strains were studied, of which 1,288 were resistant to clarithromycin. The pooled prevalence of CRHp in Africa was 27% (95% CI: 22, 33). A large discrepancy was reported among the prevalences of CRHp, ranging from 0% (95% CI: 0, 2) to 100% (95% CI: 89, 100), (I² = 95.3%, p < 0.001) (Fig. S2).

Publication bias

The presence of publication bias on the studies of CRHp in Africa was reported with a bias coefficient of 2.18 (Fig. S3).

Evolution of the prevalence of CRHp over time

Meta-regression of the prevalences of CRHp isolated in Africa over the 26 years did not reveal any significant variation ($R^2 = 0.0001$, p = 0.92), with an insignificant downward trend (slope coefficient of -0.05x).

Mutations within the 23S rRNA conferring clarithromycin resistance

Thirty studies (46%) identified ten types of mutations within the 23S rRNA (Table S1). A2143G (carried by 465 *H. pylori* strains out of 1178 tested) and A2142G (344 *H. pylori* strains out of 1027) were by far the most reported 23S rRNA mutations in Africa (p < 0.0001) (Table S1).

There was no difference between the prevalence of A2143G and A2142G in Northern Africa (p = 0.17) and southern Africa (p = 0.08). Nevertheless, the A2143G mutation was significantly more prevalent in Eastern Africa and Central Africa (Table 2). Several studies have reported cases of multiple 23S rRNA mutations in *H. pylori* strains.

Virulence genes

Eleven articles (17%) searched and reported *H. pylori* virulence genes. *cag*A (10/11; 91%) and *vac*A (6/11; 55%)

Page 4 of 13

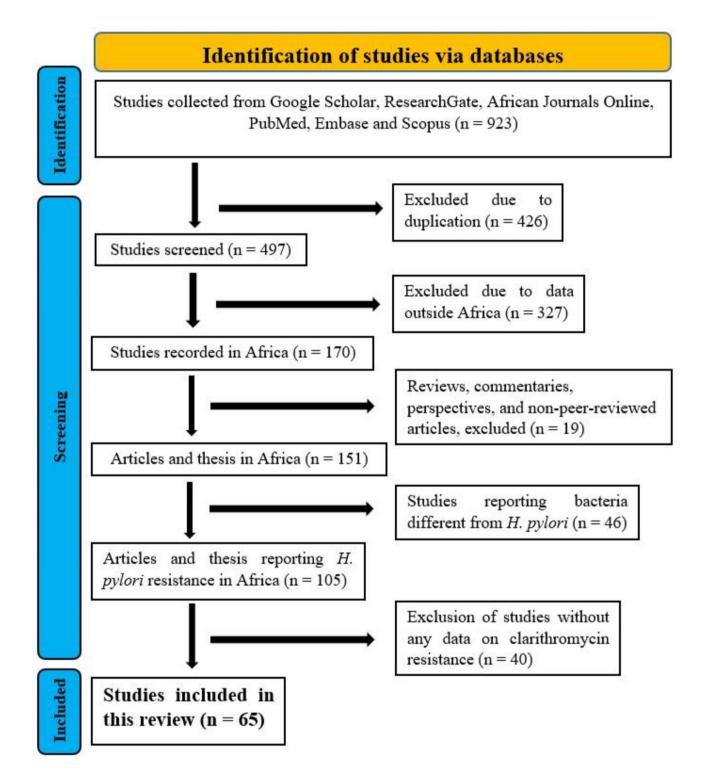


Fig. 1 PRISMA search flow diagram

à	
Ы	
st	
is s	
t	
es included in	
-	
ě	
Ы	
$\overline{\mathbf{U}}$	
.⊆∣	
ies	
÷	
stu	
S	
65	
he 6	
t	
F	
CS O	
E:	
ist	
haracteris	
IJ	
lice	
h Ú	

Country	Authors and Reference	Strains isolation period	Study design	Sample	Method used to assess clar- ithromycin resistance	Number of <i>H.</i> <i>pylori</i> strains	СКНр	%	mutations in the 235 rRNA	۶ ۸
Algeria	Bachir et al. 2018 [35]	2012-2015	NA	gastric biopsies	E-test, agar dilution method	151	38	25.2	NT	vacA, caqA,
	Djennane-Hadibi et al. 2015 [36]	2008–2014	2008-2014 prospective	gastric biopsies	Scorpion real-time PCR	91	32	35.2	A2143G (26/32; 81%), A2142G (3/32; 9.4%), A2142C (2/32; 6.3%)	ž
	Bachir et al. 2018 [37]	2014–2016 NA	NA	gastric biopsies	E-test, real time PCR	212	53	25	A2143G (69/232), A2142G (69/232)	Γ
	Raaf et al. 2017 [38]	2015–2016	2015–2016 prospective study	gastric biopsies	disc diffusion method, E-test, real-time PCR	27	0	33.3	A2143G (14/60), A2142G (14/60) NT	ΤN
Burkina Faso	Sia et al. 2018 [39]	2017	prospective	gastric biopsies	Scorpion real time PCR	132	12	9.1	A2143G (12/12)	LΝ
Cameroon	Kouitche Mabeku et al. 2019 [40]	2013-2015		gastric biopsies	Kirby–Bauer disc diffusion method	140	19	13.6	NT	ΓN
	Ndip et al. 2008 [41]	2006	NA	gastric biopsies	disc diffusion method	132	59	44.7	NT	LΝ
Chad	Bessimbaye et al. 2021 [42]	2020-2021	observational diagnostic study	gastric biopsie, stool	disc diffusion method	59	10	16.9	ИТ	ΓN
Republic of the Congo	Ontsira-Ngoyi et al. 2015 [43]	2013-2014	cross-sectional	gastric biopsies	real-time PCR	56	-	1.8	A2142G (1/1) A2143G (1/1)	ΓN
Ivory Coast	Diplo et al. 2017 [44]	2015-2016 NA	NA	gastric biopsies	Classic PCR	98	26	26.5	NA	LΝ
Democratic Republic of Congo	Tshibangu-Kabamba et al. 2020 [45]	2017–2018	NA	gastric biopsies	agar dilution method	102	24	23.5	A2142G (4/24; 16.7%), A2143G (17/24; 70.8%)	LZ
Ethiopia	Erkihun et al. 2023 [46]	2019	cross- sectional	gastric biopsies	disc diffusion method	24	16	66.7	ИТ	ΝΤ
	Asrat et al. 2004 [47]	NA	NA	NA	E-test	50	0	0	NT	LΝ
Egypt	Elrakeeb et al. 2021 [48]	2019–2020	2019–2020 prospective ran- طمستعط فلينظر	gastric biopsies	disc diffusion assay	52	17	32.7	NT	ΝT

_
0
-
c u u
=
_
~
_
-
_
~
~
0
()
0
\sim
_
—
A 1
w.
_
-
-0
_
8

Table 1 (continued)	ntinued)									
Country	Authors and Reference	Strains isolation period	Study design	Sample	Method used to assess clar- ithromycin resistance	Number of H. <i>pylori</i> strains	СКНр	%	mutations in the 23S rRNA	۶۸
Egypt	El Sayed Zaki et al. 2016 [49]	2014–2015	NA	gastric biopsies	disc diffusion assay, PCR-RFLP	72	36	50	A2143G (15/72), A2142G (13.9%; 10/72)	NT
	Ramzy et al. 2016 [50]	2013	NA	gastric biopsies	PCR-RFLP	70	40	57.1	A2142G 39/70), A2143G (1/70)	NT
	Hussien et al. 2022 [34]	2018-2019	NA	gastric biopsies	agar dilution method	19	18	94.7	NT	NT
	Sherif et al. 2004 [5 1]	2002-2003	NA	gastric biopsies	E-test	48	2	4.2	NT	NT
	Abdallah et al. 2023 [52]	NA	cross-sectional	gastric biopsies	disc diffusion method	30	15	50	NT	NT
	Eshra et al. 2023 [53]	2022	cross-sectional	gastric biopsies	E-test	40	21	52.5	NT	NT
	Hanafy and Seleem, 2019 [54]	2016–2018	observational study	gastric biopsies	E-test, RFLP-PCR	49	12	24.5	A2142G (9/12), A2143G (3/12)	NT
	Abd El Azeem et al. 2017 [55]	NA	NA	stool	disc diffusion method	23	1	47.8	NT	<i>cag</i> A, <i>bab7</i> allele of <i>bab</i> A2
	Attia et al. 2022 [56]	NA	NA	gastric biopsies	real-time PCR	32	12	37.5	A2142C (4/12), A2143G (12/32)	NT
	Hamza et al. 2018 [57]	NA	NA	stool	PCR-RFLP, disc diffusion method	20	12	60	A2142G (2/12), A2143G (3/12)	cagA, vacA
	Labeeb and El-khyat, 2019 [58]	NA	cross- sectional	gastric biopsies	agar dilution method	18	6	50	NT	cagA
	Abdelsami et al. 2020 [59] 2019–2020 NA	2019-2020	NA	gastric biopsies	E-test, RFLP-PCR	50	27	54	A2142G (5/50), A2143G (21/50)	NT
	Soltan et al. 2018 [60]	2016-2017	cross-sectional	gastric biopsies	RFLP-PCR	71	47	66.2	A2142G (39/71), A2143G (12/71)	NT
	El-Gazzar et al. 2020 [61]	2019–2020	prospective obser- vational study	gastric biopsies	disc diffusion method	28	6	32.1	NT	NT
	El Sayed Zaki et al. 2016 [62]	2015	NA	gastric biopsies	disc diffusion method	69	49	71	NT	ΤN
	Ghaith et al. 2016 [63]	2013	cross-sectional study	gastric biopsies	RFLP-PCR	70	39	55.7	55.7 A2142G (39/70)	NT

 CONTINUED	(D) D

Country	Authors and Reference	Strains isolation period	Study design	Sample	Method used to assess clar- ithromycin resistance	Number of <i>H.</i> <i>pylori</i> strains	СКНр	%	mutations in the 23S rRNA	5
Egypt	Fathi et al. 2013 [64]	2011-2012	cross-sectional study	gastric biopsies	disc diffusion method, E-test	16	16	100	ИТ	ureC,
	Elzaher et al. 2022 [65]	2018-2021	prospective study	Gastric biopsies	real-time PCR	20	S	25	NA	NA
	Metwally et al. 2022 [66]	2018–2020	cross-sectional study	Gastric biopsies	disc diffusion method	20	8	40	TN	NT
	Diab et al. 2018 [67]	2015-2017	NA	gastric biopsies	RT-PCR	60	4	6.7	A2143G (4/60)	NT
	Mahmoud et al. 2018 [68]	2016–2017	NA	gastric biopsies	disc diffusion method	70	48	68.6	L.	cagA, iceA1, vacA s1, vacA s2,
	Awad et al. 2020 [69]	ΑN	cross-sectional study	gastric biopsies	NA	30	15	50	LΝ	NT
	Ghazy et al. 2022 [<mark>70</mark>]	NA	NA	gastric biopsies	RT-PCR	42	27	64.3	23S mutant	cagA
Gambia	Secka et al. 2013 [71]	NA	NA	gastric biopsies	agar dilution	64	0	0	NT	NT
Kenya	Kimangʻa et al. 2010 [<mark>72</mark>]	NA	NA	gastric biopsies	E-test	65	0	0	NT	NT
	Lwai-Lume et al. 2005 [73]	2003–2004	cross-sectional descriptive study	gastric biopsies	NA	166	11	6.6	TN	NT
	Kabuthi et al. 2021 [74]	2018–2019	cross-sectional descriptive study	gastric biopsies	E-test	68	6	13.2	NT	NT
Morocco	Bouihat et al. 2016 [75]	2015-2016	prospective	gastric biopsies	E-test, real time scorpion PCR	177	45	25.4	A2142G (29/51; 56.9%); A2143G (20/51; 39.2%) A2142C (4/51; 7.8%)	μN
	Essaidi et al. 2022 [76]	2017-2020	NA	gastric biopsies	PCR - RFLP	96	14	14.6	A2143G	NT
Mozambique	lsmail et al. 2023 [77]	2017-2020	cross-sectional descriptive	gastric biopsies	Standard PCR and sequencing	96	10	10.4	A2142G (2/10), A2143G (8/10)	NT
Mauritania	Khiddi et al. 2020 [78]	2018	NA	gastric biopsies	Real time PCR	76	4	5.3	A2143G (4/4)	cagA
Nigeria	Oyedeji et al. 2009 [79]	AN	NA	gastric biopsies	disc diffusion method, E-test, PCR-RFLP, sequencing	186	0	0	NF	NT
Nigeria	Aboderin et al. 2007 [80]	2002-2003 2005-2006	prospective	gastric biopsies	disc diffusion assay	32	32	100	ΝΤ	NT
	Bello et al. 2019 [81]	2011-2013	cross-sectional	gastric biopsies	disc diffusion method	109	0	0	NT	NT
	Harrison et al. 2017 [82]	2010-2013	cross-sectional	gastric biopsies	E-test, PCR, sequencing	111	16	14.4	A2143G (1/16), A2144G (2/16), A2143C (2/16), C2196T	cagA, vacA s1/m1
	Ani et al. 1999 [83]	1997-1998	NA	tissue biopsies	E-test	55	7	12.7	NA	NT
	Adeniyi et al. 2012 [84]	NA	NA	gastric biopsies	disc diffusion method	43	c	7	NT	NT
	Palamides et al. 2020* [85]	2015-2018	NA	gastric biopsies	E-test	88	23	26.1	NT	cagA, vacA

Country	Authors and Reference	Strains isolation period	Study design	Sample	Method used to assess clar- ithromycin resistance	Number of H. pylori strains	· CRHp	%	mutations in the 235 rRNA	БУ
Reunion Island	Zemali et al. 2016 [86]	2014	NA	gastric biopsies	real-time PCR	73	6	12.3	A2142C (1/9); A2142G/A2143G (8/9)	NT
Senegal	Seck et al. 2013 [87]	2007–2009	AN	gastric biopsies	E-test, scorpion PCR	108	. 	0.9	A2143G (1/1)	LΝ
South Africa	Palamides et al. 2020* [85]	2015-2018	AN	gastric biopsies	E-test	132	22	16.7	ЛТ	cagA, vacA
	Tanih et al. 2010 [88]	NA	NA	gastric biopsies	disc diffusion method, agar dilution method	200	40	20	NA	NT
	Tanih and Ndip, 2013 [89]	NA	AA	gastric biopsies	disc diffusion method, Geno- Type HelicoDR kit	78	12	15.4	A2147G (12/17; 70.6%); A2146C (1/17; 5.9%)	NT
	Tanih et al. 2011 [90]	NA	NA	gastric biopsies	disc diffusion method, end-point PCR, sequencing	200	40	20	A2142G (3/3), A2143G (1/3)	NT
Sudan	Albasha et al. 2021 [91]	2018-2019	NA	gastric biopsies	PCR and DNA sequencing	25	6	36	A2142G (1/9), A2143G (5/9), T2182C (4/9), C2195T (3/9)	NT
Tanzania	Jaka et al. 2019 [92]	2014-2016	2014–2016 cross-sectional study	gastric biopsies	real-time PCR	188	54	28.7	A2143G (32/54), A2142G (22/54), A2142C (1/54), A2143C (1/54)	NT
Tunisia	Ben Mansour et al. 2010 [93]	2005-2007	prospective multi- centre study	gastric biopsies	E-test, scorpion real-time PCR	273	42	14.6	A2143G (37/42; 88.1%); A2142G (5/42; 11.9%)	NT
	Ben Mansour et al. 2016 [94]	2009	prospective study	gastric biopsies	E-test	21	ŝ	14.3	NT	NT
	Chtourou et al. 2022 [95]	2017-2020	2017–2020 cross-sectional study	gastric biopsies	Allplex <i>H. pylori</i> , ClariR PCR Assay	95	30	31.6	A2143G (90.5%; 86/95), A2142G (11,6%; 11/95)	NT
Uganda	Angol et al. 2017 [96]	2012-2013	cross-sectional study	stool	GenoType HelicoDR PCR	21	9	28.6	NT	NT
Zambia	Kebotsamang et al. 2024 [97]	ЧZ	case-control study	gastric biopsies	Bosphore <i>Helicobacter</i> <i>pylori</i> Genotyping Kit v1 (real- time PCR)	174	48	27.6	A2143G (38/48), A2142G (29/48)	NT

(2025) 14:31

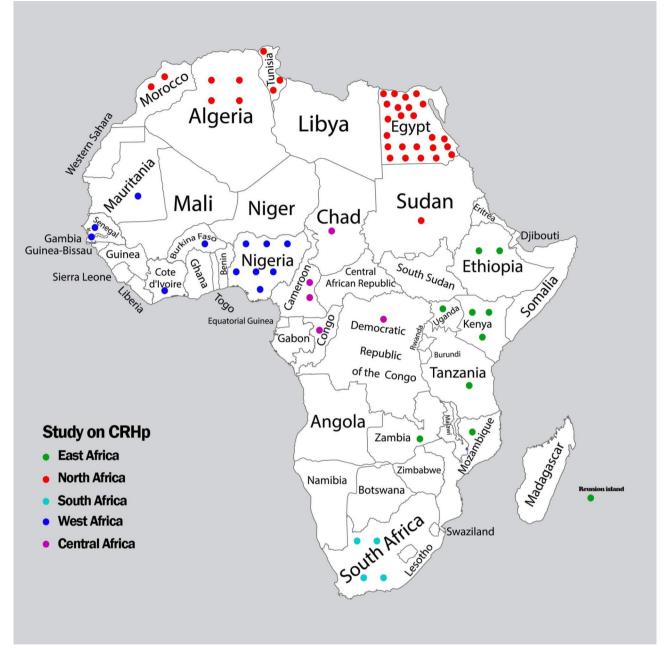


Fig. 2 Map of Africa showing the number of study on CRHp in Africa

Table 2	Distribution of A2143G and A2142G mutations	in
African r	gions	

Africa and regions	Mutation within	n the 23S rRNA	<i>p</i> -value
	A2143G n/N (%)	A2142G n/N (%)	_
Northern Africa	342/996 (34)	275/878 (31)	0.17
Western Africa	18/33 (55)	0	-
Eastern Africa	86/121 (71)	61/121 (50)	0.001*
Central Africa	18/25 (72)	5/25 (20)	0.0002*
Southern Africa	1/3 (33)	3/3 (100)	0.08
Africa	465/1178 (40)	344/1027 (34)	0.003*

*: significant difference (*p* < 0.05)

were the most reported, followed by *ice*A1 (1/11; 9%), *ure*C (1/11; 9%) and *bab*A2 (1/11; 9%).

Discussion

In the reviewed studies, the agar dilution method, considered the gold standard for antimicrobial susceptibility testing (AST) for *H. pylori*, was used in only 6 of the 65 studies included. In contrast, the Kirby-Bauer disc diffusion method, which is regarded as an unreliable AST method for detecting CRHp, remains the most commonly used method on the African continent (in 22 studies out of 65). This underscores the urgent need for implementing standardized microbiological methods for resistance detection to enhance the accuracy and validity of surveillance data.

The overall pooled prevalence of CRHp reported in this study was 27% (95% CI: 22, 33), and was very similar to the global pooled prevalence of CRHp from 248 articles in 2023 [27.53% (95% CI: 25.41, 29.69)] [20]. The pooled prevalence of CRHp observed in Africa in this systematic review and meta-analysis was slightly lower than that reported for the Asian continent (29.57% and 35.97% in 2023) [85, 86]. However, the CRHp prevalence in Africa was slightly higher than that (22%) reported in the Asia-Pacific region [21], and was the same as that (27%) reported for the Southern Asian region in 2023 [22]. Concerning Europe, the African CRHp pooled prevalence was quite similar to those reported for the European continent, 25% in 2021 [23] and 26.25% in 2023 [20]. The pooled prevalence of CRHp reported in this study was higher than that (23.86%) observed in the American continent in 2023 [20], and was significantly higher than the 10.3% [24] observed for Oceania in 2020 and the 7.02% reported for North America in 2023 [25].

In this study, we reported a steady trend ($R^2 = 0.0001$; p = 0.92) for the prevalence of CRHp in Africa for strains isolated from 1997 to 2022, with a slightly negative slope coefficient of -0.05x. This is contrary to most of the trends observed worldwide. An increase from 24.28% in 2010-2017 to 32.14% in 2018-2021 was reported in a global study that included 248 articles in 2023 [20]. In addition, increases in CRHp prevalence were also observed in the South-East Asian Region (13% in 2006-2008 to 21% in 2012–2016) [9], South Asia (21% in 2003–2012 to 30% in 2013–2022) [22], Oceania (6.4% in 1997–2000 to 16.1% in 2000-2013) [24]. Similar to this study, steady trends of CRHp have been reported for Latin America (11% in 1996-2000, 12% in 2001-2005, 14% in 2006-2011, p = 0.47) [26]. Furthermore, a decreasing trend in CRHp prevalence was noted for the European continent (25% in 2013-2016 to 20% (2017-2020), p = 0.002) [23].

This systematic review and meta-analysis reported ten types of *H. pylori* 23S rRNA mutations conferring clarithromycin resistance in Africa. All the reported 23S rRNA mutations have been already observed worldwide [27–29]. Our results are similar to those reported in Iran: A2143G (46.6%), A2142G (37.2%), and A2142C (5.5%) [30]. However, other 23S rRNA mutations conferring clarithromycin resistance in *H. pylori*, such as A2146G, C2772T, C2759T, G2212A, and A2144G, reported in Brazil [31], Mexico [27], Portugal [32], South Korea [33], and Iraq [34] have not been observed in Africa.

Conclusion

The data reported in this systematic review and metaanalysis showed that the pooled prevalence of CRHp in Africa was very similar to the overall pooled prevalence observed globally. In addition, more representative studies on *H. pylori* are needed in African countries. This can help to appreciate the real extent of antibiotic resistance in *H. pylori* strains. While waiting for further representative studies of the African population to be carried out, the treatment of *H. pylori* infections must be based on the guidelines of the AHMSG first Lagos consensus, for which triple therapy was suggested as this could be tenable.

Supplementary Information

The online version contains supplementary material available at https://doi.or g/10.1186/s13756-025-01533-6.

Supplementary Material 1

Supplementary Material 2

Author contributions

K.M.D. conceptualized and designed the study, helped in data acquisition and analysis, and drafted the manuscript. T.B. helped in data acquisition and analysis. S.S. helped in data analysis. L.L., B.S.B., A.S., and S.D. substantively revised the manuscript. E.E.I., F.P.S. and M.C. provided writing assistance and substantively revised the manuscript. A.E.K. supervised the study and substantively revised the manuscript. S.I.S. supervised the study, and substantively revised the manuscript.

Funding

Not applicable.

Data availability

Data is provided within the manuscript or supplementary information files.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Author details

¹Department of Microbiology, Global Health Research Institute, Lomé, Togo

²Laboratoire des Sciences Biomédicales Alimentaires et de Santé Environnementale, Ecole Supérieure des Techniques Biologiques et Alimentaires, Université de Lomé, Lomé, Togo

³Ecole Doctorale des Sciences Juridiques, Politiques, Economiques et de Gestion, Cheikh Anta Diop University, Dakar, Senegal

⁴Medical Microbiology division, Medical Laboratory Services, University of Benin Teaching Hospital, Benin City, Nigeria

⁵State Key Laboratory for Animal Disease Control and Prevention, Guangdong Laboratory for Lingnan Modern Agriculture, College of Veterinary Medicine, South China Agricultural University,

Guangzhou 510642, China

⁶World Health Organization Country Office Senegal, Dakar, Senegal ⁷Faculté de Médecine, Pharmacie et Odontostomatologie, Cheikh Anta Diop University, Dakar, Senegal ⁸Fundamental Sciences Department, Health Sciences Faculty, Université de Kara, Kara, Togo

⁹Department of Internal Medicine, School of Veterinary Medicine and Animal Science, University of São Paulo, São Paulo, Brazil

¹⁰School of Veterinary Medicine, Metropolitan University of Santos, Santos, Brazil

¹¹Bacteriology-Virology laboratory, National University Teaching Hospital Aristide Le Dantec, Dakar, Senegal

¹²Department of Botany and Microbiology, Faculty of Science, Suez Canal University, Ismailia 41522, Egypt

¹³Department of Biology, College of Science, Imam Muhammad Ibn Saud Islamic University (IMSIU), Riyadh 11623, Saudi Arabia

¹⁴Molecular Biology and Biotechnology Department, Nigerian Institute of Medical Research, Yaba, Lagos, Nigeria

¹⁵Department of Biological Sciences, Mountain Top University, Prayer, Ogun State, Nigeria

Received: 10 October 2024 / Accepted: 15 February 2025 Published online: 12 April 2025

References

- Hooi JKY, Lai WY, Ng WK, Suen MMY, Underwood FE, Tanyingoh D, et al. Global prevalence of *Helicobacter pylori* infection: systematic review and Meta-analysis. Gastroenterology. 2017;153:420–9.
- Chen Y-C, Malfertheiner P, Yu H-T, Kuo C-L, Chang Y-Y, Meng F-T, et al. Global prevalence of *Helicobacter pylori* infection and incidence of gastric Cancer between 1980 and 2022. Gastroenterology. 2024;166:605–19.
- Smith SI, Schulz C, Ugiagbe R, Ndip R, Dieye Y, Leja M, et al. *Helicobacter* pylori Diagnosis and treatment in Africa: The First Lagos Consensus Statement of the African *Helicobacter* and Microbiota Study Group. Dig Dis. 2024;42:240–56.
- Segal I, Ally R, Mitchell H. *Helicobacter pylori*—an African perspective. QJM: Int J Med. 2001;94:561–5.
- Moss SF. The clinical evidence linking *Helicobacter pylori* to Gastric Cancer. Cell Mol Gastroenterol Hepatol. 2017;3:183–91.
- de Martel C, Georges D, Bray F, Ferlay J, Clifford GM. Global burden of cancer attributable to infections in 2018: a worldwide incidence analysis. Lancet Glob Health. 2020;8:e180–90.
- Cellini L, Donelli G. Virulence factors of *Helicobacter pylori*. Microb Ecol Health Disease. 2000;12:259–62.
- Baj J, Forma A, Sitarz M, Portincasa P, Garruti G, Krasowska D, et al. *Helicobacter* pylori Virulencefactors - mechanisms of bacterial pathogenicity in the gastric microenvironment. Cells. 2020;10:27
- Savoldi A, Carrara E, Graham DY, Conti M, Tacconelli E. Prevalence of Antibiotic Resistance in *Helicobacter pylori*: a systematic review and Meta-analysis in World Health Organization regions. Gastroenterology. 2018;155:1372–e138217.
- 10. Hsieh PF, Yang JC, Lin JT, Wang JT. Molecular mechanisms of clarithromycin resistance in *Helicobacter pylori*. J Formos Med Assoc. 1998;97:445–52.
- Alarcón-Millán J, Fernández-Tilapa G, Cortés-Malagón EM, Castañón-Sánchez CA, De Sampedro-Reyes J, Cruz-del Carmen I, et al. Clarithromycin resistance and prevalence of *Helicobacter pylori* virulent genotypes in patients from Southern México with chronic gastritis. Infect Genet Evol. 2016;44:190–8.
- Lin Y, Shao Y, Yan J, Ye G. Antibiotic resistance in *Helicobacter pylori*: from potential biomolecular mechanisms to clinical practice. J Clin Lab Anal. 2023;37:e24885.
- Malfertheiner P, Megraud F, Rokkas T, Gisbert JP, Liou J-M, Schulz C et al. Management of *Helicobacter pylori* infection: the Maastricht VI/Florence consensus report. Gut. 2022;71:gutjnl–202
- World Health Organization. WHO publishes list of bacteria for which new antibiotics are urgently needed. 2017. https://www.who.int/news/item/27-0 2-2017-who-publishes-list-of-bacteria-for-which-new-antibiotics-are-urgentl y-needed. Accessed 10 Feb 2023.
- Jaka H, Rhee JAh, Östlundh L, Smart L, Peck R, Mueller A, et al. The magnitude of antibiotic resistance to *Helicobacter pylori* in Africa and identified mutations which confer resistance to antibiotics: systematic review and metaanalysis. BMC Infect Dis. 2018;18:193.
- Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ. 2021;372:n71.

- Munn Z, Moola S, Riitano D, Lisy K. The development of a critical appraisal tool for use in systematic reviews addressing questions of prevalence. Int J Health Policy Manag. 2014;3:123–8.
- Egger M, Smith GD, Phillips AN. Meta-analysis: principles and procedures. BMJ. 1997;315:1533–7.
- Duval S, Tweedie R. Trim and fill: a simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. Biometrics. 2000;56:455–63.
- Sholeh M, Khoshnood S, Azimi T, Mohamadi J, Kaviar VH, Hashemian M, et al. The prevalence of clarithromycin-resistant *Helicobacter pylori* isolates: a systematic review and meta-analysis. PeerJ. 2023;11:e15121.
- Hong T-C, El-Omar EM, Kuo Y-T, Wu J-Y, Chen M-J, Chen C-C, et al. Primary antibiotic resistance of *Helicobacter pylori* in the Asia-Pacific region between 1990 and 2022: an updated systematic review and meta-analysis. Lancet Gastroenterol Hepatol. 2024;9:56–67.
- 22. Shrestha AB, Pokharel P, Sapkota UH, Shrestha S, Mohamed SA, Khanal S, et al. Drug resistance patterns of commonly used antibiotics for the Treatment of *Helicobacter pylori* Infection among south Asian countries: a systematic review and Meta-analysis. Trop Med Infect Disease. 2023;8:172.
- Bujanda L, Nyssen OP, Vaira D, Saracino IM, Fiorini G, Lerang F, et al. Antibiotic resistance prevalence and trends in patients infected with *Helicobacter pylori* in the period 2013–2020: results of the European Registry on *H. Pylori* Management (Hp-EuReg). Antibiotics. 2021;10:1058.
- Schubert JP, Gehlert J, Rayner CK, Roberts-Thomson IC, Costello S, Mangoni AA, et al. Antibiotic resistance of *Helicobacter pylori* in Australia and New Zealand: a systematic review and meta-analysis. J Gastroenterol Hepatol. 2021;36:1450–6.
- Khoshnood S, Pakzad R, Kaviar VH, Hashemian M, Karamollahi S, Sadeghifard N et al. Global Estimate of Clarithromycin Resistance in Clinical isolates of *Helicobacter pylori*: a systematic review and Meta-analysis. Clin Lab. 2023;69.
- 26. Camargo CM, García A, Riquelme A, Otero W, Camargo CA, Hernandez-García T, et al. The Problem of *Helicobacter pylori* Resistance to antibiotics: a systematic review in Latin America. Official J Am Coll Gastroenterol| ACG. 2014;109:485.
- Camorlinga-Ponce M, Gómez-Delgado A, Aguilar-Zamora E, Torres RC, Giono-Cerezo S, Escobar-Ogaz A et al. Phenotypic and Genotypic Antibiotic Resistance Patterns in *Helicobacter pylori* strains from ethnically Diverse Population in México. Front Cell Infect Microbiol. 2021;10.
- Seo J-H, Woo H-O, Youn H-S, Rhee K-H. Antibiotics resistance of *Helicobacter* pylori and treatment modalities in children with *H. Pylori* infection. Korean J Pediatr. 2014;57:67–71.
- Stone GG, Shortridge D, Flamm RK, Versalovic J, Beyer J, Idler K, et al. Identification of a 23S rRNA gene mutation in Clarithromycin-Resistant *Helicobacter pylori*. Helicobacter. 1996;1:227–8.
- Khademi F, Sahebkar A. An updated systematic review and Meta-analysis on the *Helicobacter pylori* Antibiotic Resistance in Iran (2010–2020). Microb Drug Resist. 2020;26:1186–94.
- de Costa ACM SF da, Cahú AKM, de Sena AR, Leite AF, de Leite AB, Sampaio TCC et al. Epidemiological study of antibiotic-resistant *Helicobacter pylori* in Brazil a meta-analysis. Caderno Pedagógico. 2024;21:e4568–e4568.
- Marques AT, Vítor JMB, Santos A, Oleastro M, Vale FF. Trends in *Helicobacter* pylori resistance to clarithromycin: from phenotypic to genomic approaches. Microb Genomics. 2020;6.
- Gong EJ, Ahn JY, Kim JM, Lee SM, Na HK, Lee JH, et al. Genotypic and phenotypic resistance to Clarithromycin in *Helicobacter pylori* strains. J Clin Med. 2020;9:1930.
- Hussein RA, Al-Ouqaili MTS, Majeed YH. Detection of clarithromycin resistance and 23SrRNA point mutations in clinical isolates of *Helicobacter pylori* isolates: phenotypic and molecular methods. Saudi J Biol Sci. 2022;29:513–20.
- Bachir M, Allem R, Tifrit A, Medjekane M, Drici AE-M, Diaf M, et al. Primary antibiotic resistance and its relationship with cagA and vacA genes in *Helicobacter pylori* isolates from Algerian patients. Braz J Microbiol. 2018;49:544–51.
- Djennane-Hadibi F, Bachtarzi M, Layaida K, Ali Arous N, Nakmouche M, Saadi B, et al. High-level primary clarithromycin resistance of *Helicobacter pylori* in Algiers, Algeria: a prospective Multicenter Molecular Study. Microb Drug Resist. 2016;22:223–6.
- Bachir M, Allem R, Benejat L, Tifrit A, Medjekane M, Drici AE-M, et al. Molecular detection of mutations involved in *Helicobacter pylori* antibiotic resistance in Algeria. J Antimicrob Chemother. 2018;73:2034–8.
- Raaf N, Amhis W, Saoula H, Abid A, Nakmouche M, Balamane A, et al. Prevalence, antibiotic resistance, and MLST typing of *Helicobacter pylori* in Algiers. Algeria Helicobacter. 2017;22:e12446.

- Sia R, Plouzeau C, Godonou H, Guingane A, Coulibaly A, Somda S et al. Lowlevel primary clarithromycin resistance of *Helicobacter pylori* in Burkina Faso: a prospective molecular study. Helicobacter. 2018;24.
- 40. Kouitcheu Mabeku LB, Eyoum Bille B, Tepap Zemnou C, Tali Nguefack LD, Leundji H. Broad spectrum resistance in *Helicobacter pylori* isolated from gastric biopsies of patients with dyspepsia in Cameroon and efflux-mediated multiresistance detection in MDR isolates. BMC Infect Dis. 2019;19:880.
- Ndip RN, Malange Takang AE, Ojongokpoko JEA, Luma HN, Malongue A, Akoachere J-FTK, et al. *Helicobacter pylori* isolates recovered from gastric biopsies of patients with gastro-duodenal pathologies in Cameroon: current status of antibiogram. Tropical Med Int Health. 2008;13:848–54.
- Bessimbaye N, Moussa AM, Habkréo M, Moukhtar AS, Ouchemi C. Biochemical and resistance profile of *Helicobacter pylori* isolated in N'Djamena in Chad. J Drug Delivery Ther. 2021;11:33–41.
- 43. Ontsira Ngoyi EN, Atipo Ibara BI, Moyen R, Ahoui Apendi PC, Ibara JR, Obengui O, et al. Molecular detection of *Helicobacter pylori* and its Antimicrobial Resistance in Brazzaville, Congo. Helicobacter. 2015;20:316–20.
- Bernadette D, Valérie G, Nathalie G, Francis Y, Solange K, Aboulaye O, et al. Molecular detection of Antibiotic Resistance of *Helicobacter pylori* from gastric biopsies in Abidjan (Côte d'Ivoire). Microbiol Res J Int. 2017;20:1–7.
- 45. Tshibangu-Kabamba E, Ngoma-Kisoko P, de Tuan J, Matsumoto VP, Akada T, Kido J. Next-generation sequencing of the whole bacterial genome for Tracking Molecular Insight into the Broad-Spectrum Antimicrobial Resistance of *Helicobacter pylori* Clinical isolates from the Democratic Republic of Congo. Microorganisms. 2020;8:887.
- Erkihun M, Chanie DN, Siraj YA. Antimicrobial-Resistance Profile of *Helico-bacter pylori*, obtained from endoscopic patients in Bahir Dar, North West Ethiopia. Can J Infect Dis Med Microbiol. 2023;2023;7326288.
- Asrat D, Kassa E, Mengistu Y, Nilsson I, Wadström T. Antimicrobial susceptibility pattern of *Helicobacter pylori* strains isolated from adult dyspeptic patients in Tikur Anbassa University Hospital, Addis Ababa, Ethiopia. Ethiop Med J. 2004;42:79–85.
- Elrakeeb A, Eltayyeb SH, Elgazzar A, Elgendy AA, Alboraie M. Tailored Helicobacter pylori therapy is more effective than conventional therapy: a randomized-controlled trial. Al-Azhar Assiut Med J. 2021;19:379.
- Sayed Zaki ME, Sherif DM, Abdelwahab Ali M, Shehta A, Megahed A, Latif Alsayed MA, et al. Molecular Study of Clarithromycin Resistant *Helicobacter pylori* strains from Egyptian centre. IntJCurrMicrobiolAppSci. 2016;5:165–73.
- Ramzy I, Elgarem H, Hamza I, Ghaith D, Elbaz T, Elhosary W, et al. Genetic mutations affecting the first line eradication therapy of *Helicobacter pylori*infected Egyptian patients. Rev Inst Med trop S Paulo. 2016;58:88.
- Sherif M, Mohran Z, Fathy H, Rockabrand DM, Rozmajzl PJ, Frenck RW. Universal High-Level Primary Metronidazole Resistance in *Helicobacter pylori* isolated from children in Egypt. J Clin Microbiol. 2004;42:4832–4.
- Abdallah NMA, Morsy RME, Awad YMM, Fathy MS, El Deeb MT. *Helicobacter pylori* antibiotic resistance patterns among Egyptian children and predictors of resistance. Microbes Infect Dis. 2023;4:1287–95.
- 53. Eshra K, Amer I, El Sharaby R, El Sharawy S, Eissa R. Detection of *hef*A gene in multidrug resistant *Helicobacter pylori* at Tanta University Hospital. Microbes Infect Dis. 2023;4:514–21.
- 54. Hanafy AS, Seleem WM. Refractory *Helicobacter pylori* gastritis: the hidden predictors of resistance. J Glob Antimicrob Resist. 2019;19:194–200.
- Azeem AE, Abdel-Ghaffar ME, Hassan Shokaeir BA-R, Ali M. Genotyping of Helicobacter pylori isolates from Egyptian patients. Int J Biosci. 2017;10:121–8.
- Attia NM, Elsilimy H, Khalil AF, Baddour NM, El Sawaf GEDA, Shawky SM. Detection of A2142G, A2142C and A2143G clarithromycin mutations in *Helicobacter pylori* in Alexandria University Pediatric Hospital. Microbes Infect Dis. 2022;3:434–41.
- 57. Hamza D, Elhelw R, Elhariri M, Ragab E. Genotyping and antimicrobial resistance patterns of *Helicobacter pylori* in human and dogs associated with A2142G and A2143G point mutations in clarithromycin resistance. Microb Pathoq. 2018;123:330–8.
- Labeeb AZ, EL-Khyat AH. Relation between *Helicobacter pylori cag*A gene Status and Antibiotic Resistance Pattern in Peptic Ulcer patients. Egypt J Med Microbiol. 2019;28:69–76.
- Abdelsami SA, Fari AA, Abbas SI, Elgazzar AA. Determination of Antibiotic Resistance and Relevance of 23S rRNA gene in *Helicobacter pylori* with Clarithromycin Resistance in Chronic Gastritis. Benha J Appl Sci. 2020;5(2):41–7. Issue 8 part (1)-(.
- Soltan MA, Mansour MA, Zahir TI, Rashed HE, Fahmy YA. Prediction of *Helico-bacter pylori* Clarithromycin Resistance by detection of point mutations in 23S rRNA gene. Egypt J Med Microbiol. 2018;27:13–20.

- 61. El-Gazzar A, Al-Boraie M, Hanem El-Tayyeb S, El-Rakeeb A. Empirical versus susceptibility-based eradication therapy for *Helicobacter pylori* infection in Egypt. Al-Azhar Med J. 2020;49:1931–42.
- 62. Zaki MES, Othman W, Ali MA, Shehta A, Fluoroquinolone-Resistant. *Heli-cobacter pylori* strains Isolated from One Egyptian University Hospital: Molecular Aspects. 2016;:6.
- 63. Ghaith D, Elzahry M, Mostafa G, Mostafa S, Elsherif R, Ramzy I. Mutations affecting domain V of the 23S rRNA gene in *Helicobacter pylori* from Cairo. Egypt J Chemother. 2016;28:367–70.
- 64. Fathi MS, EL-Folly RF, Hassan RA, El-Arab ME. Genotypic and phenotypic patterns of antimicrobial susceptibility of *Helicobacter pylori* strains among Egyptian patients. Egypt J Med Hum Genet. 2013;14:235–46.
- Elzaher EA, EL-Defrawy MS, Elhosiny NF, Behairy OG. Detection of Antimicrobial Resistance genes in children suffering from *Helicobacter pylori* infection. Benha J Appl Sci. 2022;7:179–86.
- Metwally M, Ragab R, Abdel Hamid HS, Emara N, Elkholy H. *Helicobacter pylori* Antibiotic Resistance in EaysingleScenterCstudy Study. Infect Drug Resist. 2022;15:5905–13.
- 67. Diab M, El-Shenawy A, El-Ghannam M, Salem D, Abdelnasser M, Shaheen M, et al. Detection of antimicrobial resistance genes of *Helicobacter pylori* strains to clarithromycin, metronidazole, Amoxicillin and tetracycline among Egyptian patients. Egypt J Med Hum Genet. 2018;19:417–23.
- Mahmoud AB, Makled AF, Abdoo AG, El Shayeb ASI, El Askary SA, Sleem AS. Different detection methods of Virulent *Helicobacter pylori* in gastric biopsies. Egypt J Med Microbiol. 2018;27:109–18.
- Awad YMMM, Eldeeb MT, Fathi MS, Mahmoud N, Morsy RME. Helicobacter pylori Antibiotic resistance patterns among Egyptian children and predictors of resistance. QJM: Int J Med. 2020;113 Supplement_1:hcaa063.021.
- 70. Ghazy A, Haydara T, Osman EM, Elhadidy A. Relevance of Multi-drug resistant *H. pylori* in recurrent gastritis. Int Med J. 2022;29:30.
- Secka O, Berg DE, Antonio M, Corrah T, Tapgun M, Walton R, et al. Antimicrobial susceptibility and resistance patterns among *Helicobacter pylori* strains from the Gambia, West Africa. Antimicrob Agents Chemother. 2013;57:1231–7.
- Kimang'a AN, Revathi G, Kariuki S, Sayed S, Devani S. *Helicobacter pylori*: prevalence and antibiotic susceptibility among kenyans. South Afr Med J. 2010;100.
- Lwai-Lume L, Ogutu EO, Amayo EO, Kariuki S. Drug susceptibility pattern of *Helicobacter pylori* in patients with dyspepsia at the Kenyatta National Hospital, Nairobi. East Afr Med J. 2005;82:603–8.
- 74. Kabuthi AN. Antimicrobial Susceptibility Profile of *Helicobacter pylori* Isolated From Patients With Dyspepsia in Two Tertiary Hospitals in Nairobi, Kenya. Thesis. UON. 2021.
- Bouihat N, Burucoa C, Benkirane A, Seddik H, Sentissi S, Al Bouzidi A, et al. *Helicobacter pylori* Primary Antibiotic Resistance in 2015 in Moracphenotypic cotypigenotypicoprospective and Multicenter Study. Microb Drug Resist. 2017;23:727–32.
- Essaidi I, Bounder G, Jouimyi RM, Boura H, Elyounsi I, Kheir F-Z, et al. Comparative study of *Helicobacter pylori* Resistance to Clarithromycin and Metronidazole and its association with Epidemiological Factors in a Moroccan Population. Asian Pac J Cancer Prev. 2022;23:2755–61.
- Ismail M, Majaliwa ND, Vale FF, Cumbana R, Sumbana JJ, Muchongo A, et al. Molecular detection of *Helicobacter pylori* and its genotypic antimicrobial resistance patterns in dyspeptic Mozambican patients. Helicobacter. 2023;28:e13000.
- Khiddi F, Abdellahi MVM, Horma MA, Billoet A, Collobert G, Amar AM, et al. Characteristics of *Helicobacter pylori* strains isolated from Mauritanian patients. Helicobacter. 2020;25:e12726.
- 79. Oyedeji KS, Smith SI, Coker AO, Arigbabu AO. Antibiotic susceptibility patterns in *Helicobacter pylori* strains from patients with upper gastrointestinal pathology in western Nigeria. Br J Biomed Sci. 2009;66:10–3.
- Aboderin OA, Abdu A, Odetoyin BW, Okeke IN, Lawa OO, Ndububa DA et al. Antibiotic resistance of *Helicobacter pylori* from patients in Ile-Ife, South-west, Nigeria. Afr Health Sci. 2007;7.
- Bello AK, Tukur AD, Yakasai AM, Borodo MM. *Helicobacter pylori* antibiotic sensitivity pattern in dyspeptic patients in Kano, Nigeria. South Afr J Infect Dis. 2019;34:1–7.
- Harrison U, Fowora MA, Seriki AT, Loell E, Mueller S, Ugo-Ijeh M, et al. *Helicobacter pylori* strains from a Nigerian cohort show divergent antibiotic resistance rates and a uniform pathogenicity profile. PLoS ONE. 2017;12:e0176454.

- Ani AE, Malu AO, Onah JA, Queiroz DMM, Kirschner G, Rocha GA. Antimicrobial susceptibility test of *Helicobacter pylori* isolated from Jos. Nigeria Trans Royal Soc Trop Med Hygiene. 1999;93:659–61.
- Adeniyi BA, Lawal TO, Otegbayo JA, Oluwasola OA, Odaibo GN, Ola SO, et al. Cultural characteristics and antibiotic susceptibility pattern of *Helicobacter pylori* isolated from Dyspepsia patients. Gastroenterol Insights. 2012;4:e21.
- Palamides P, Jolaiya T, Idowu A, Loell E, Onyekwere C, Ugiagbe R, et al. *Helico-bacter pylori* patient isolates from South Africa and Nigeria differ in virulence factor pathogenicity profile and associated gastric disease outcome. Sci Rep. 2020;10:11409.
- Zemali N, Guillemot G, Jaubert J, Picot S, Thomas E, Becquart JP, et al. *Helico-bacter pylori* resistance to clarithromycin in Reunion Island. Méd Mal Infect. 2016;46:385–9.
- Seck A, Burucoa C, Dia D, Mbengue M, Onambele M, Raymond J, et al. Primary antibiotic resistance and associated mechanisms in *Helicobacter pylori* isolates from Senegalese patients. Ann Clin Microbiol Antimicrob. 2013;12:3.
- Tanih NF, Okeleye BJ, Naidoo N, Clarke AM, Mkwetshana N, Green E, et al. Marked susceptibility of South African *Helicobacter pylori* strains to ciprofloxacin and Amoxicillin: clinical implications: original article. South Afr Med J. 2010;100:45–8.
- Tanih NF, Ndip RN. Molecular Detection of Antibiotic Resistance in South African isolates of *Helicobacter pylori*. Gastroenterol Res Pract. 2013;2013:e259457.
- 90. Tanih NF, Ndip LM, Ndip RN. Characterisation of the genes encoding resistance to metronidazole (*rdx*A and *frx*A) and clarithromycin (the 23S-rRNA genes) in South African isolates of *Helicobacter pylori*. Annals Trop Med Parasitol. 2011;105:251–9.
- 91. Albasha AM, Elnosh MM, Osman EH, Zeinalabdin DM, Fadl AAM, Ali MA, et al. *Helicobacter pylori* 23S rRNA gene A2142G, A2143G, T2182C, and C2195T

mutations associated with clarithromycin resistance detected in Sudanese patients. BMC Microbiol. 2021;21:38.

- Jaka H, Rüttgerodt N, Bohne W, Mueller A, Gross U, Kasang C, et al. *Helico-bacter pylori* Mutation conferring resistance to fluoroquinolones and Clarithromycin among dyspeptic patients attending a Tertiary Hospital, Tanzania. Can J Gastroenterol Hepatol. 2019;2019:e8481375.
- Ben Mansour K, Burucoa C, Zribi M, Masmoudi A, Karoui S, Kallel L, et al. Primary resistance to clarithromycin, metronidazole and Amoxicillin of *Helicobacter pylori* isolated from Tunisian patients with peptic ulcers and gastritis: a prospective multicentre study. Ann Clin Microbiol Antimicrob. 2010;9:22.
- 94. Ben Mansour K, Fendri C, Battikh H, Garnier M, Zribi M, Jlizi A, et al. Multiple and mixed *Helicobacter pylori* infections: comparison of two epidemiological situations in Tunisia and France. Infect Genet Evol. 2016;37:43–8.
- Chtourou L, Moalla M, Mnif B, Smaoui H, Gdoura H, Boudabous M, et al. Prevalence of *Helicobacter pylori* resistance to clarithromycin in Tunisia. J Med Microbiol. 2022;71:001561.
- Angol DC, Ocama P, Ayazika Kirabo T, Okeng A, Najjingo I, Bwanga F. Helicobacter pylori from Peptic Ulcer patients in Uganda is highly resistant to Clarithromycin and fluoroquinolones: results of the GenoType HelicoDR Test Directly Applied on Stool. Biomed Res Int. 2017;2017:e5430723.
- 97. Kebotsamang T, Munkombwe D, Bwalya L, Kelly P, Kayamba V. Prevalence of Clarithromycin-Resistant *Helicobacter pylori* strains in Zambia: a sub-saharan African Country. Dig Dis. 2024;42:1–7.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.